

CORRESPONDENCE

Open Access



# *KRAS* mutation detection by liquid biopsy for pancreatic ductal adenocarcinoma

Mahmoud Yousef<sup>1</sup>, Abdelrahman Yousef<sup>1</sup>, Mark W. Hurd<sup>2</sup>, Ashwathy Pillai<sup>3</sup>, Saikat Chowdhury<sup>1</sup>, Rebecca Snyder<sup>4</sup>, Mark Knaf<sup>5</sup>, Ryan L. Lewis<sup>6</sup>, Paul M. Roy<sup>6</sup>, Mohammad Fanaeian<sup>1</sup>, Sali Albarouki<sup>7</sup>, Luca F. Castelnovo<sup>1</sup>, Jennifer Peterson<sup>1</sup>, Brandon G. Smaglo<sup>1</sup>, Robert A. Wolff<sup>1</sup>, Shubham Pant<sup>1</sup>, Jason Willis<sup>1</sup>, Ryan Huey<sup>1</sup>, Michael Overman<sup>1</sup>, Ching-Wei Tzeng<sup>4</sup>, Michael P. Kim<sup>4</sup>, Naruhiko Ikoma<sup>4</sup>, Jess E. Maxwell<sup>4</sup>, Matthew H. G. Katz<sup>4</sup>, Huamin Wang<sup>8</sup>, Anirban Maitra<sup>2,8</sup>, Eugene Koay<sup>9</sup>, Ethan B. Ludmir<sup>9,10</sup>, Anthony Chen<sup>1</sup>, Camila Lopez<sup>1</sup>, Haoqiang Ying<sup>11</sup>, John Paul Shen<sup>1</sup> and Dan Zhao<sup>1\*</sup>

## Abstract

The clinical utility of liquid biopsy (LB) for pancreatic ductal adenocarcinoma (PDAC) remain understudied. Our single-institution cohort of 311 PDAC patients with non-tumor tissues informed LB found 81.2% positivity ( $N=186$ ) in metastatic cases and in 52.4% ( $N=43$ ) of localized disease. *KRAS* mutations were detected in 64.6% ( $N=148$ ) of metastatic cases and 16% ( $N=13$ ) for localized disease. Positive LB, especially *KRAS* mutation detection, is associated with worse overall survival (OS) in metastatic PDAC (median 14.5 vs. 31.3 months,  $HR=2.7$ ,  $95\%CI=1.7-4.3$ ,  $P<0.0001$ ). The positive concordance rates of *KRAS* and *TP53* mutations were 63% and 68% in metastatic disease but only 7% (*KRAS*) and 33% (*TP53*) in localized disease, respectively. Among the 41 patients who underwent serial liquid biopsy testing, 25% tested positive after an initial negative result. LB detects therapeutically targetable mutations in 58.5% of PDAC patients and is associated with OS.

**Keywords** Pancreatic cancer, Liquid biopsy, *KRAS*, PDAC, Molecular profiling, Mutation, OS

\*Correspondence:

Dan Zhao

dzhao3@mdanderson.org

<sup>1</sup>Department of Gastrointestinal Medical Oncology, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

<sup>2</sup>Sheikh Ahmed Center for Pancreatic Cancer Research, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

<sup>3</sup>Department of Hospital Medicine, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

<sup>4</sup>Department of Surgical Oncology, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

<sup>5</sup>Department of Genomic Medicine, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

<sup>6</sup>Department of Enterprise Data Engineering and Analytics, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

<sup>7</sup>Department of Gastroenterology and Hepatology, Baylor College of Medicine, Houston, TX, USA

<sup>8</sup>Department of Anatomical Pathology, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

<sup>9</sup>Department of Gastrointestinal Radiation Oncology, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

<sup>10</sup>Department of Biostatistics, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

<sup>11</sup>Department of Molecular and Cellular Oncology, Division of Basic Science Research, The University of Texas MD Anderson Cancer Center, Houston, TX, USA



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

**To the editor**

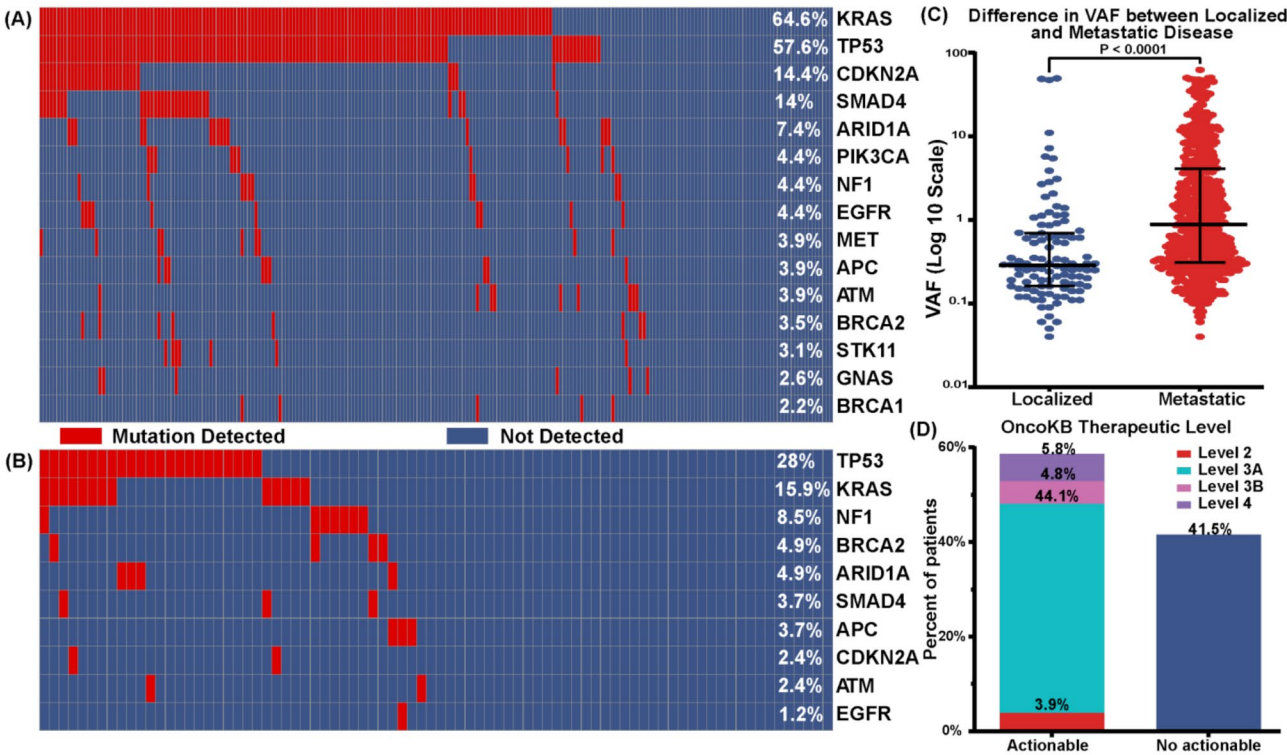
*KRAS* is mutated in approximately 90% of pancreatic ductal adenocarcinoma (PDAC) including 35% *KRAS*<sup>G12D</sup>, 30% *KRAS*<sup>G12V</sup>, 15% *KRAS*<sup>G12R</sup>, and 1-2% *KRAS*<sup>G12C</sup> [1, 2]. *KRAS*<sup>G12C</sup> inhibitors showed efficacy in PDAC and many *KRAS* inhibitors are in clinical development [3–8]. We previously reported the *KRAS* mutation by tissue testing with PDAC outcome which is associated with worse overall survival (OS) [9]. The utility of liquid biopsy (LB) is promising in PDAC [10, 11]. CtDNA positive rate was 29.48% by tumor-informed whole exome sequencing (WES) in post-surgical PDAC patients on surveillance [12]. There are few real-world data on the non-tumor tissue informed liquid biopsy testing.

**Results**

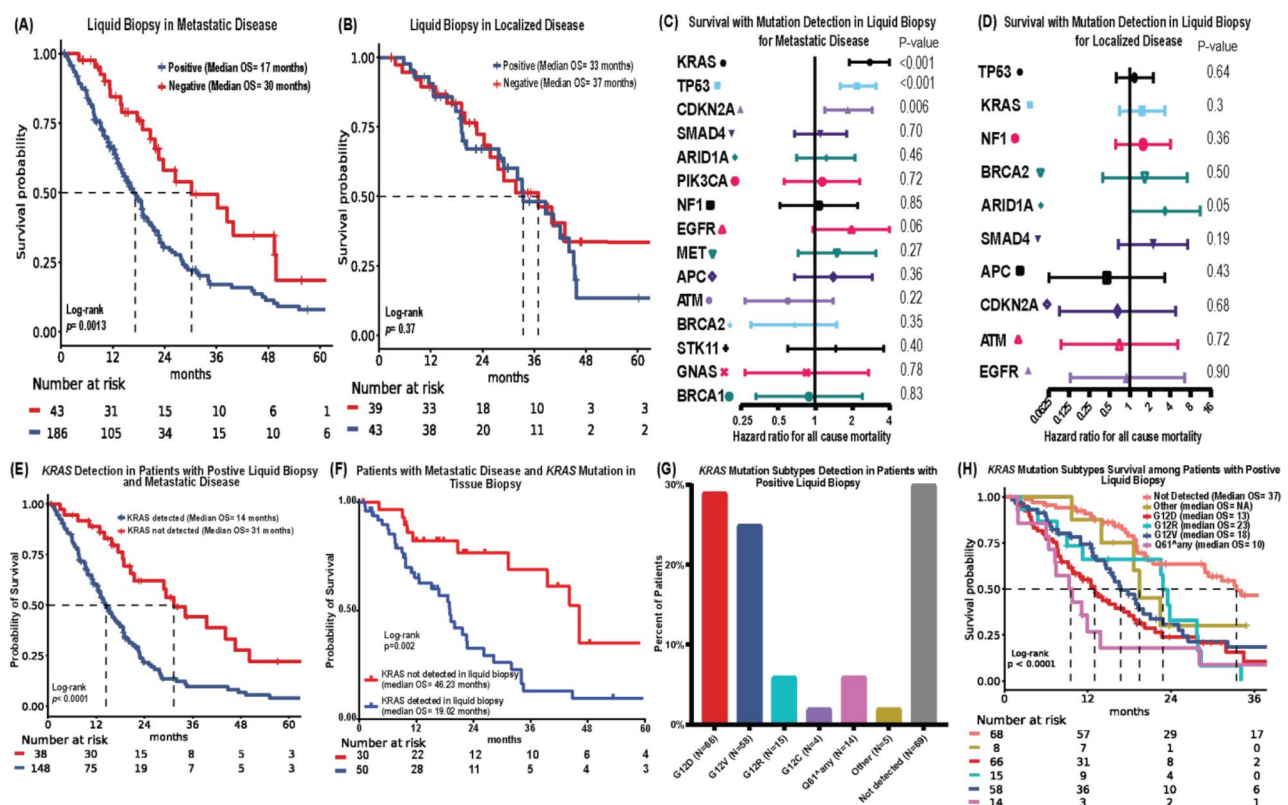
We analyzed 311 PDAC patients underwent in-house non tumor informed ctDNA testing from 2018 to 2023 at MD Anderson cancer center. 73% (*N*=229) had metastatic disease (Supplemental Methods, Table S1). The median follow-up was 34.9 months with median OS 22.5 months (95% CI=19.2–25.8). The median age at diagnosis was 64.9 years old. LB was positive in 81.2% (*N*=186) of metastatic cases 52.4% (*N*=43) of localized disease. *KRAS* mutations were detected in 64.6% (*N*=148) metastatic disease, followed by *TP53* (57.6%, *N*=132, Fig. 1-A).

However, for localized disease, the most detected mutation was *TP53* (28%, *N*=23), followed by *KRAS* (16%, *N*=13) (Fig. 1-B). Median VAF in localized disease was significantly lower than metastatic disease, medians (interquartile range)=0.29 (0.53) vs. 0.88 (3.78) respectively, *P*<0.001, Fig. 1-C). LB detected actionable mutations in 58.5% (*N*=182) of all patients tested according to the OncoKB therapeutic level of evidence classification, with 3.9% (*N*=12) at level 2, 44.1% (*N*=137) at level 3 A, 4.8% (*N*=15) at level 3 B, and 5.8% (*N*=18) at level 4 (Fig. 1-D). The positive concordance rate for the subset of patients underwent tissue biopsy testing (*n*=116), was 63% (*n*=50/80) for *KRAS* mutation, 68% (*n*=43/63) for *TP53* mutation, 26% (*n*=5/19) for *SMAD4*, and 80% (*n*=8/10) for *CDKN2A* in metastatic disease. Localized disease had lower positive concordance rate, with 7% (*n*=2/27) for *KRAS* and 33% (*n*=7/21) for *TP53* (Table S2-3).

Positive LB was associated with worse OS (HR=2.1, 95%CI=1.3–3.3, *P*=0.0015) in metastatic disease (Fig. 2A). The OS difference was not significant (HR=1.3, 95%CI=0.72–2.5, *P*=0.36; Fig. 2B) in localized disease. Univariate COX regression analyses for OS in metastatic cases showed that mutations in *KRAS* (HR=2.8, 95%CI=1.9-4, *P*<0.001), *TP53* (HR=2.19, 95%CI=1.6–3.1, *P*<0.001), and *CDKN2A* (HR=1.85, 95%CI=1.2–2.9,



**Fig. 1** Mutations detected in LB and OS. **A-** Oncoplot for mutations detected in metastatic disease at LB. **B-** Oncoplot for mutations detected in localized disease at LB. **C-** Difference in median VAF of mutations detected in LB between localized disease and metastatic disease. **D-** Rates of actionable mutations detected in LB by OncoKB therapeutic levels



**Fig. 2** Outcomes with positivity of LB and with *KRAS* mutations detection. **A**- OS with positive LB in metastatic disease. **B**- OS with positive LB in localized disease. **C**- Hazard ratios (HRs) of OS with mutation detection by LB in metastatic disease. **D**- HRs of OS with mutation detection in LB in localized disease. **E**- OS of patients with positive *KRAS* mutation vs. other mutations in LB for metastatic disease. **F**- OS of metastatic disease patients with positive *KRAS* mutation detected by tissue NGS stratified by *KRAS* mutation detection in LB. **G**- Frequencies of detected *KRAS* mutation subtypes. **H**- OS of patients with *KRAS* mutation detected in LB by *KRAS* mutation subtypes

$P=0.006$ ) were associated with worse OS (Fig. 2-C-D). *KRAS* mutation detection in LB for metastatic disease was associated with worse OS (median 14.5 vs. 31.3 months, HR=2.7, 95%CI=1.7–4.3,  $P<0.001$ ; Fig. 2-E) but the OS difference was not significant in localized disease (Figure S1-A). Notably, in metastatic cases with *KRAS* mutation detected by tumor tissue testing, *KRAS* detection in LB was associated with worse OS (HR=2.57, 95%CI=1.42–4.63,  $P=0.002$ ; Fig. 2-F). The most frequent *KRAS* mutation detected was *KRAS*<sup>G12D</sup> ( $N=66$ , 41%), followed by *KRAS*<sup>G12V</sup> ( $N=58$ , 36%, Fig. 2-G). *KRAS*<sup>G12D</sup> and *KRAS*<sup>Q61</sup> detection was associated with poorer OS in patients with positive liquid biopsy (Fig. 2-H), which is consistent with our previous findings in patients who had tissues testing [9].

Among 41 patients who underwent multiple LB tests, 25% were initially ctDNA-negative then subsequently tested positive. None of the 22 patients with positive results converted to negative in the subsequent tests. Among 35 patients who received systemic treatment, patients with increased number of detected mutations ( $n=16$ ) had a trend of worse OS (median OS 22.9 months vs. 26.4 months) compared with decreased number of

mutations ( $n=3$ ; HR=2.1, 95%CI=0.48–15.02,  $P=0.37$ , Figure S1-B). Patients with increased VAF for *KRAS* ( $n=18$ , median OS=18.7 months) or *TP53* ( $n=13$ , median OS=22.9 months) showed a tendency towards worse OS compared to patients with decreased VAF of *KRAS* ( $n=8$ , median OS=44.8 months; HR=2.02, 95%CI=0.73–5.59,  $P=0.18$ , Figure S2-A-C) or decreased VAF of *TP53* ( $n=4$ , median OS=34 months; HR=1.95, 95%CI=0.54–7.04,  $P=0.31$ , Figure S2-D-F).

## Conclusion

We found that 81.2% ( $N=186$ ) were LB positive in patients with metastatic disease and 52.4% ( $N=43$ ) positivity rate in localized disease of PDAC. *KRAS* mutations were detected in 64.6% ( $N=148$ ) of patients with metastatic disease, while only 16% ( $N=13$ ) of patients had localized disease (Fig. 1). The detection of any mutation in LB was associated with worse OS in metastatic PDAC (Fig. 2). Moreover, *KRAS* mutations, especially *KRAS*<sup>G12D</sup> and *KRAS*<sup>Q61</sup>, were associated with worse OS (Fig. 2).

## Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s13045-025-01696-0>.

Supplementary Material 1

## Acknowledgements

This work was supported by The University of Texas MD Anderson Cancer Center Context Engine and the Context Engine Team. The Context Engine is MD Anderson's institutional Data Management System and Digital Architecture.

## Author contributions

D.Z. and M.Y. conceptualized the paper, data interpretation, and writing the manuscript. D. Z., A.Y., A. P., M.Y., S.C., J.P.S. conceived and designed the study, contributed to literature search, data acquisition, data analysis, data interpretation, and manuscript writing. S.C., M.K. R.L., P.R., M. F., S.A. contributed to data acquisition, data analysis, data interpretation. R.S., B.S., R.W., S.P., J.W., R.H., M.O., C.W.T., M.K., N.I., J.M., M.K., E.K., E.L. contributed to patient enrollment, treatment, assessment, and data interpretation. H.W., H.Y. and A.M. contributed to data acquisition and data interpretation. M.K., L.C., S.C., J.P., A. C., C.L. and E.K. contributed to the statistical analysis and writing of the manuscript.

## Funding

This work was supported by the Col. Daniel Connelly Memorial Fund, the National Cancer Institute (K22 CA234406 to J.P.S., and the Cancer Center Support Grant (P30 CA016672), the Cancer Prevention & Research Institute of Texas (RR180035 & RP240392 to J.P.S., J.P.S. is a CPRIT Scholar in Cancer Research), the Appendiceal Cancer Pseudomyxoma Peritonei Research Foundation, and the Conquer Cancer Career Development Award to J.P.S. Role of the Funder/Sponsor: The funders had no role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, or approval of the manuscript; or decision to submit the manuscript for publication. Disclaimer: Any opinions, findings, and conclusions expressed in this material are those of the author(s) and do not necessarily reflect those of the American Society of Clinical Oncology or Conquer Cancer.

## Data availability

Individual patient-level data are not publicly available to maintain compliance with HIPAA regulations and IRB protocol. Anonymized data are available for non-commercial use from the corresponding author upon request, pending data usage agreement and/or IRB-approved collaboration.

## Declarations

## Ethics approval and consent to participate

This study was approved by the MD Anderson Institutional Review Board (IRB), protocol number 2023-0091. A waiver of informed consent was granted per the USA federal regulation 45 CFR 46.116(f) for this retrospective study.

## Competing interests

Brandon Smaglo: Consulting for Ipsen. Shubham Pant: Advisory for Zymeworks, Ipsen, Novartis, Janssen, Boehringer Ingelheim, and AskGene Pharma; and he receives research funding from Mirati Therapeutics, Lilly, Xencor, Novartis, Rgenix, Bristol-Myers Squibb, Astellas Pharma, Framewave, 4D Pharma, Boehringer Ingelheim, NGM Biopharmaceuticals, Janssen, Arcus Biosciences, Elicio Therapeutics, Bionte, Ipsen, Zymeworks, Pfizer, ImmunoMET, Imuneering, and Amal Therapeutics. Anirban Maitra: Consultant for Tezcat Biosciences is listed as an inventor of a patent licensed to Thrive Earlier Detection (an Exact Sciences Company) relevant to early detection of pancreatic cancer. John Paul Shen: Grant/research support/collaboration: Celsius Therapeutics, BostonGene, Caris Life Sciences, Natera, Xilis, Palantir,

Genentech. Consulting/stock ownership: Egen Biosciences, NaDeNo Nanoscience. Dan Zhao: Clinical trials with hMirati/BMS, Phanes, CARsgen, TriSalus and Affini-T; Consulting for Ipsen.

Received: 19 February 2025 / Accepted: 2 April 2025

Published online: 17 April 2025

## References

1. Cancer Genome Atlas Research Network. Electronic address Aadhe, cancer genome atlas research N: integrated genomic characterization of pancreatic ductal adenocarcinoma. *Cancer Cell*. 2017;32(2):185–203. e113.
2. Bailey P, Chang DK, Nones K, Johns AL, Patch AM, Gingras MC, Miller DK, Christ AN, Bruxner TJ, Quinn MC, et al. Genomic analyses identify molecular subtypes of pancreatic cancer. *Nature*. 2016;531(7592):47–52.
3. Canon J, Rex K, Saiki AY, Mohr C, Cooke K, Bagal D, Gaida K, Holt T, Knutson CG, Koppada N, et al. The clinical KRAS(G12C) inhibitor AMG 510 drives anti-tumour immunity. *Nature*. 2019;575(7781):217–23.
4. Hallin J, Engstrom LD, Hargis L, Calinisan A, Aranda R, Briere DM, Sudhakar N, Bowcut V, Baer BR, Ballard JA, et al. The KRAS <sup>G12C</sup> Inhibitor MRTX849 provides insight toward therapeutic susceptibility of KRAS-Mutant cancers in mouse models and patients. *Cancer Discov*. 2020;10(1):54–71.
5. Hong DS, Fakih MG, Strickler JH, Desai J, Durm GA, Shapiro GI, Falchook GS, Price TJ, Sacher A, Denlinger CS, et al. KRAS(G12C) Inhibition with Sotorasib in advanced solid tumors. *N Engl J Med*. 2020;383(13):1207–17.
6. Kemp SB, Cheng N, Markosyan N, Sor R, Kim IK, Hallin J, Shoush J, Quinones L, Brown NV, Bassett JB, et al. Efficacy of a Small-Molecule inhibitor of KrasG12D in immunocompetent models of pancreatic cancer. *Cancer Discov*. 2023;13(2):298–311.
7. Arbour KC, Puneekar S, Garrido-Laguna I, Hong DS, Wolpin B, Pelster MS, Barve M, Starodub A, Sommerhalder D, Chang S, et al. 6520 preliminary clinical activity of RMC-6236, a first-in-class, RAS-selective, tri-complex RAS-MULTI(ON) inhibitor in patients with KRAS mutant pancreatic ductal adenocarcinoma (PDAC) and non-small cell lung cancer (NSCLC). *Ann Oncol*. 2023;34:S458.
8. Jiang L, Menard M, Weller C, Wang Z, Burnett L, Aronchik I, Steele S, Flagella M, Zhao R, Evans JWW, et al. Abstract 526: RMC-9805, a first-in-class, mutant-selective, covalent and oral KRASG12D(ON) inhibitor that induces apoptosis and drives tumor regression in preclinical models of KRASG12D cancers. *Cancer Res*. 2023;83(7Supplement):526–526.
9. Yousef A, Yousef M, Chowdhury S, Abdilleh K, Knafli M, Edelkamp P, Alfaro-Munoz K, Chacko R, Peterson J, Smaglo BG, et al. Impact of KRAS mutations and co-mutations on clinical outcomes in pancreatic ductal adenocarcinoma. *Npj Precision Oncol*. 2024;8(1):27.
10. Kruger S, Heinemann V, Ross C, Diehl F, Nagel D, Ormanns S, Liebmann S, Prinz-Bravin I, Westphalen CB, Haas M, et al. Repeated MutKRAS ctDNA measurements represent a novel and promising tool for early response prediction and therapy monitoring in advanced pancreatic cancer. *Ann Oncol*. 2018;29(12):2348–55.
11. Bernard V, Kim DU, San Lucas FA, Castillo J, Allenson K, Mulu FC, Stephens BM, Huang J, Semaan A, Guerrero PA, et al. Circulating nucleic acids are associated with outcomes of patients with pancreatic cancer. *Gastroenterology*. 2019;156(1):108–e118104.
12. Botta GP, Abdelrahim M, Drengler RL, Aushev VN, Esmail A, Laliotis G, Brewer CM, George GV, Abbate SM, Chandana SR et al. Association of personalized and tumor-informed ctDNA with patient survival outcomes in pancreatic adenocarcinoma. *Oncologist* 2024.

## Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.